



L91C045

(For Professional Use Only) V4

1. Intended Purpose

The TANBead® Nucleic Acid Extraction Kit is a nucleic acid purification kit based on magnetic bead technology by using with corresponding TANBead® Nucleic Acid Extractor, which can automatically isolate and purify cell-free DNA (cfDNA) from human serum and plasma. The purified cfDNA can be used with any downstream application employing PCR-based qualitative, semi-quantitative, quantitative assays and capillary electrophoresis. The kit is intended for use by technicians, physicians, and biologists with well-trained in molecular biological techniques, the techniques of magnetic bead purification and *in vitro* diagnostic procedures. Any diagnostic results generated by using the sample preparation procedure in conjunction with any downstream diagnostic assay should be interpreted related to other clinical or laboratory findings. The kit is not limited to any specific disorder, condition, or other additional accompanying diagnostics. It is applicable for all population.

2. The basic principle

The silicon dioxide layer coated on the magnetic beads can adsorb the negatively charged molecules to purify nucleic acids from samples.

3. Specification

Starting Materials	1~6 mL plasma / serum
Elution Volume	50~100 μL
Tomic at DNIA cristal	1~100 ng per mL plasma /
Typical DNA yield	serum
Typical DNA size	100~300 bp

⚠ DNA yield will vary from 1~100 ng according to sample source and storage method. The average DNA yield is 20 ng per mL of serum from healthy individuals.

4. Component Supplied with the Kit

component	appirou mini mi	V 40			
Lysis buffer	120 mL x 4	Guanidine salt, tris buffer, surfactant			
Washing buffer 1	60 mL x 1	Guanidine salt, tris buffer, pH 8.0			
Washing buffer 2	24 mL x 1	Add 96 mL 95~100% EtOH before use			
Magnetic Beads	1.5 mL x 2	Magnetic Beads			
Elution Buffer	1.5 mL x 4	Nuclease-Free Water			
Proteinase K	1.0 mL x 12	Proteinase K, store at 2~8 °C			
Protocol	1	Instruction guide for user			

5. Auto Plate Content and Plate Position

Plate Position	Buffer	Volume (μL)
1	Lysis Buffer	As described in Chapter 11
2	Lysis Buffer	As described in Chapter 11
3	Washing Buffer 1	1000 μL
4	Washing Buffer 2	1000 μL
5	Washing Buffer 2	1000 μL
6	Elution Buffer	50~100 μL
7	N/A	N / A
8	Spin Tip	<u>-</u>

6. Kit Storage and Shelf Life

- 1) Components under room temperature (15~35°C) can be stored until the expiration date labeled on the box.
- 2) The proteinase K is transported at room temperature. Upon received, please store proteinase K at $2\sim8^{\circ}$ C.

7. Precautions

- 1) It can be used for in vitro diagnostic.
- 2) When the temperature is below 20°C, place the reagent plate in an oven (preheated 42~60°C) from 5 to 10 minutes.
- Avoid vigorous shaking, in order to avoid excessive formation of foam.

- 4) Carefully remove aluminum foil to avoid splashing.
- Do not expose the opened reagents or Auto Plates / Auto Tubes to air. The evaporation would lead to pH change, or effect on the extraction effectiveness.
- 6) The reagents are all colorless and transparent. Colored reagents indicate contamination, please replace it with a fresh Auto Plates / Auto Tubes before proceeding.
- 7) Please check the integrity of the Auto Plates / Auto Tubes and remember to mount the spin tips into the appropriate position of the suitable instrument before operating them.
- 8) Please wear a mask and disposable gloves when handling.
- 9) Use sterile consumables to avoid nuclease contamination.
- 10) Reagent solution contains guanidine salt, avoid using bleach containing detergent.
- Avoid eyes, skin, and clothing contact with reagents. In case of any contact, flush with flowing water.
- 12) If any serious incident occurs, please report to the manufacturer and the competent authority of the member state in which the user and / or the patient is established.

8. Materials required, Not Supplied

- TANBead[®] Nucleic Acid Extraction System Model: Maelstrom 2410 series (non-sterile)
- 2) Disposable gloves
- 3) Scissors, utility knives
- 4) Micropipette, disposable tips (10 μ L / 200 μ L / 1000 μ L)
- 5) 1.5 mL microcentrifuge tube
- 6) 15 mL / 50 mL conical tube
- 7) 20% SDS (Sodium dodecyl sulfate)
- 8) Streck Cell-Free DNA BCT tubes or other Cell-Free DNA collection tube

9. Sample Collection, Storage and Transportation

Sample collection and storage

1) Plasma

Σ/19

- a. Blood samples collected in Streck Cell-Free DNA BCT tubes is recommended. It is stable for up to 14 days.
- b. Centrifuge whole blood at 1600 x g for 10 minutes.
- c. Transfer the upper plasma layer to a new conical tube.
- d. Centrifuge at 16000 x g for 10 minutes.
- e. Carefully transfer the supernatant to a new conical tube for cfDNA isolation.

2) Serum

- a. Blood samples collected in serum separator tube.
- b. Let the blood clot for at least 30 minutes in room temperature.
- c. Centrifuge whole blood at 1600 x g for 10 minutes.
- d. Transfer the upper serum layer to a new conical tube.
- e. Centrifuge at 16000 x g for 10 minutes.
- f. Carefully transfer the supernatant to a new conical tube for cfDNA isolation.

3) Specimen storage

- a. After centrifugation, the plasma / serum can be stored at
 - i. 2~8°C up to 7 days.
 - ii. -20°C for long-term preservation.

■ Specimen transportation

Transportation of whole blood, plasma and serum specimens should be followed by specific pathogen transportation-related regulations. The whole blood sample should be kept between $2\sim25^{\circ}\text{C}$ during transportation and within 6 hours for separated serum. Plasma/ Serum sample can be transported between $2\sim8^{\circ}\text{C}$ or by frozen.

10. Nucleic Acids Extraction Protocol

 Mix sample with 20% SDS and proteinase K as described in below table.

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Sample volume	20% SDS	Proteinase K	Sample volume	20% SDS	Proteinase K
1 mL	60 µL	40 µL	4 mL	240 µL	160 µL
2 mL	120 µL	80 µL	5 mL	300 μL	200 μL
3 mL	180 µL	120 µL	6 mL	360 µL	240 µL

- Mix well and incubate at 60°C for 20 minutes.
- Load reagent buffers into 24-well plates as described in below table

Plate No.	Buffer	Sa	ample Volum	ne	Dises No	Buffer	Sample Volume			
Plate No.	Buller	1 mL	2 mL	Plate No.		Buller	4 mL	5 mL	6 mL	
1	LB	1600 μL	д 3200 µL 4800 µL 1 LB 3200 µL 4000 µL				4000 µL	4800 μL		
1	MB		30 µL		•	MB		30 µL		
2			2	LB	3200 μL	4000 µL	4800 μL			
					2	MB		30 µL		
3	WB1		1000 μL		3	WB1		1000 μL		
4	WB2		1000 μL		4	WB2	1000 μL			
5	WB2		1000 μL		5	WB2	1000 μL			
6	EB		50 - 100 μL		6	EB		50 - 100 μL		

4) Add sample into Auto Plate as described in below table.

Sample volume	Plate 1	Plate 2	Sample volume	Plate 1	Plate 2
1 mL	1 mL		4 mL	2 mL	2 mL
2 mL	2 mL		5 mL	2.5 mL	2.5 mL
3 mL	3 mL		6 mL	3 mL	3 mL

- 5) Select a program "CFDNA-SLB" for 1~3 mL input samples; select a program "CFDNA-DLB" for 4~6 mL input samples. The parameters are given in the following section.
- 6) Follow the guide shown on the screen and place plates carefully. Make sure that the chamfer of the plate faces toward the same direction.
- 7) Carefully remove the Plates when the program is finished.
- Use micropipette to transfer the purified nucleic acids from plate #6 to a clean tube.
- 9) Discard used Plates and spin tips into the waste recycling bin.

11. Program

■ Maelstrom 2410 series (for 1~3 mL input samples)

Program Name: CFDNA-SLB												
Plate				2		3	4	5	6	7	8	
Volume (μL)	780	00	-		1000	1000	1000	100	100) -	
Keep Ten	np.	-		-		-		-	-	-	-	
Action	1	Fo U/		-		For. U/D	For. U/D	For. U/D	For.	For		
Name		LE	3	-		WB1	WB2	WB2	EB	-	TIP	
Step	Pla	ate		mp. °C)		lixing min)	Mixing (rpm)	Collec (sec)		por nin)	Pause	
1		1	(Off		10	1000	60		0	Off	
2		3				10	1000	60		0	Off	
3	·	4		-		2	1500	30		0	Off	
4		5		-	2		1500	30	1	.0	Off	
5	6		(Off		8	1500	60		0	Off	
6		5	-		- 0.2		1500	0		0	Off	
8		6		0		0.2	1500	0		0	Off	

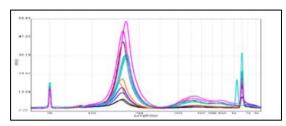
■ Maelstrom 2400 series (for 4~6 mL input samples)

Program Name: CFDNA-DLB														
Plate		1		2		3		4	5		6	7		8
Volume (μL)	780	00	780	0	1000		1000	1000	1	100	100)	-
Keep Ten	np			-		-		-	-		1	-		-
Action)	Fo U/		For U/I		For. U/D		For. U/D	For. U/D	For.		For.		-
Name		LE	3	LB		WB1		WB2	WB2		EB	-		TIP
Step	Pla	ate		mp. °C)		lixing min)		Mixing (rpm)	Collec (sec)	t	Var (m	oor in)	ı	Pause
1		1	(Off		10		1000	60		()		Off
2		2		-		10		1000	60		()		Off
3		3		-		10		1000	60		(0		Off
4		4		-		2		1500	30		()		Off
5		5		-		2		1500	30		1	0		Off

Step	Plate	Temp. (°C)	Mixing (min)	Mixing (rpm)	Collect (sec)	Vapor (min)	Pause
6	6	Off	8	1500	60	0	Off
7	5	-	0.2	1500	0	0	Off

12. Result

Total DNA yield was quantified using Qubit dsDNA HS assay kit: 1~100 ng per mL serum / plasma. The expected peak bp (100~300 bp fragments) and low amount of gDNA contamination were observed by using Qsep 100 capillary gel electrophoresis system. The cfDNA isolate from 1-6 mL serum was shown below.



13. Reagent performance

■ Repeatability

Under repeatability conditions where nucleic acids are extracted with the same reagent kit on the same source samples by the same operator. The coefficient of variation of nucleic acid extraction concentration is less than 5%.

Reproducibility

A five-day reproducibility test was carried out with the same source samples for 5 consecutive days with the same reagent kit by different operators. The coefficient of variation of nucleic acid extraction concentration is less than 5%.

■ The stability of extracted DNA

Storage Conditions	DNAstability				
-80°C	Over 90 days				
-20°C	28 days				
4°C	14 days				
25°C	2 days				
Freeze-thaw	5 times				

14. Explanation of Symbols

***	Manufacturer	Ţį	Consult instructions for use
15°C 35°C	Temperature limit	Σ	Contains sufficient for test
C€	CE mark	IVD	<i>In vitro</i> diagnostic medical use
REF	Catalogue number	\triangle	Caution
LOT	Batch code	NON	Non-sterile
2	Do not re-use	*	Keep away from sunlight
w	Date of manufacture	8	Use-by date

EC REF

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15. Post-market surveillance conclusion

After a risk assessment and clinical evaluation assessment, when weighing the benefits of medical device, patients, and the risks associated with the use of the device, the risk is acceptable. The postmarket surveillance report shows that no death or serious adverse events occurred.

